FORMULASI DAN EVALUASI MIKROSFER Ca-ALGINAT DENGAN MODEL ANTIGEN OVALBUMIN DENGAN TEKNIK AEROSOLISASI

FORMULATION AND EVALUATION OF Ca-ALGINATE MICROSPHERES ENTRAPPING MODEL ANTIGEN PRODUCED BY AEROSOLIZATION TECHNIQUE

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Abstract

Ovalbumin protein was generally used as a model of vaccine antigen for oral delivery system. This research highlighted the advantages of Ca-alginate microspheres entrapping ovalbumin which prepared by ionotropic gelation using aerosolisation technique. It involved alginate as polymer and calcium chloride as crosslinker. The objective of this study was to investigate the preparation and characterization of microspheres produced from CaCl₂ with different concentration and alginate in terms of size, protein loading, entrapment efficiency, and yield. The alginate concentrations used in this study were 1.5% and 2.5% w/v. While, the CaCl₂ concentrations used in this study were 0.1, 0.5 and 1.5 M. The results showed that low concentration of CaCl₂ could not produce microspheres, instead it produced gelling sheet. Microspheres were formed by increasing the CaCl₂ concentration. Infra red spectra were also showed that crosslinked between alginate molecule and Ca²⁺ was occurred. Smaller microspheres was produced by increasing CaCl₂ concentration from 0,5 M to 1,5 M. The effect of

increasing alginate concentration from 1.5% to 2.5% w/v showed an increase of protein loading, entrapment efficiency, and yield. From particle size examination, increasing alginate concentration showed a decrease of particle size. In summary, high entrapment efficiency, high protein loading and microspheres yield, and small particle size of ovalbumin-alginate microsphere were successfully produced using the ionotropic gelation-aerosolization method which can be utilized as one of oral protein or vaccine delivery systems.

Keywords: Microspheres, Ovalbumin, Alginate, characterization, Aerosolization.

Introduction

Ovalbumin is egg white glycoprotein that comprises 385 aminoacids (molecular weight 43 kDa) (1). It is easily denatured at high temperature and acid pH (1). Administering oral antigen is the most effective way to induce immunological tolerance to protein antigens (2). Thus, ovalbumin as an antigen, could stimulate the formation of antibodies and improve immunity. Polymer microspheres have been investigated to protect antigen from acid pH and enzymatic degradation in gastrointestinal tract. Therefore, it may sustain or control the release of the antigen vaccine (3).

Umer *et al* (4) defined microencapsulation as a process to entrap materials using polymer form microspheres from 1 to 1000 micron. In general, microencapsulation techniques consist of chemical and mechanical methods. Current study applies ionotropic gelation method based on polyelectrolyte capability to form hydrogel using polymer and crosslinking agent. Aerosolization technique was used because it is a cost effective, fast, simple technique. Moreover, it does not involve organic solvent which can contribute to protein integrity (5).

Polymer is required to coat drug or the core of active substance (6). Sodium alginate is a biodegradable and biocompatible natural polymer, non toxic to the body, cheap and most commonly used as polymer in the microparticles (7).

Crosslinking agents are usually cations such as Pb²⁺, Cd²⁺, Zn²⁺, Cu²⁺, Co²⁺, Ca²⁺, Ba²⁺, dan Sr²⁺ (8). Calcium ions have been extensively used as crosslinking agents due to low toxicity and ability to form a good gel and provided significant stability (9).

Several factors affect the microparticles preparation such as concentration of polymer and crosslinking agents (10). Higher polymer concentration produced bigger microspheres, but more spherical in shape (11). Crosslinking agents also influenced particle size. Lower crosslinking agents, produced fragile and amorphous microspheres, even it could not form the microspheres (12). Higher concentration of crosslinker produced smaller microspheres size as a result of stronger binding between them, but often resulted rough surface (10). Therefore, this research was conducted to study the formulation and characterization of Ca-alginate microspheres entrapping ovalbumin using different concentration of alginate polymer and CaCl₂ crosslinker.

Materials and Methods

Materials

Ovalbumin *pharmaceutical grade*; Natrium alginate *pharmaceutical grade*; CaCl₂.2H₂O *pharmaceutical grade*; Sodium citrate *pharmaceutical grade*; NaCl p.a; NaH₂PO₄ p.a; KH₂PO₄ p.a; HCl p.a; NaOH p.a; *protein quantification kit*, Aquadest.

Methods

Ca-Alginate microsphere preparation using Ionotropic Gelation – Aerosolization

Preparation of Ca-alginate microsphere using ionotropic gelation method by aerosolization techniques could be explained as follows: Alginate solution (concentration of 1.5 and 2.5%) containing 2.5% ovalbumin was sprayed into crosslinking agent CaCl₂ solution (concentration of 0.1, 0.5 and 1.5M) and was stirred continuously for 2 hours at 1000 rpm. The microspheres were collected by

centrifugation at 2500 rpm for 6 minutes, washed two times with aquadest and finally

freeze	Alginate concentration (%)		CaCl ₂ concentration (M)
dried 20	2.5	1.5	
•	B1	A1	0.1
hours at -	B2	A2	0.5
	В3	A3	1.5
80°C.			

Ca-Alginate microspheres formulations were summarized in **Table 1**.

TABLE 1. Ovalbumin-alginate microspheres formulation

A1: Alginate 1.5% and $CaCl_20.1$ M; A2: Alginate 1.5% and $CaCl_20.5$ M A3: Alginate 1.5% and $CaCl_21.5$ M; B1: Alginate 2.5% and $CaCl_20.1$ M B2: Alginate 2.5% and $CaCl_20.5$ M; B3: Alginate 2.5% and $CaCl_21.5$ M

Protein Loading

Loading of ovalbumin into microspheres was analysed following breakdown of 400 mg of microspheres suspensions in 50 mL sodium citrate solution over 12 hours at 1000 rpm at room temperature. The drug content was determined using protein quantification assay using UV spectrophotometry at 600 nm.

Encapsulation efficiency and Yield

Encapsulation efficiency was determined as equation below:

Encapsulation efficiency (%) =
$$\underline{\text{amount of ovalbumin}}$$
 $x = 100$

Theoretical amount of ovalbumin

Yield (%) = $\underline{\text{weight of microspheres}}$ $x = 100$

total weight of polymer and protein

Results and Discussion

Almost spherical and some rough morphological microspheres were observed. However, microspheres with 0.1M CaCl₂ produced gelling sheets. Some rough surface was caused by no cryoprotectant agent to protect microspheres during freeze drying such as sucrose and lactose.

From evaluation of infra red spectra we can see that wavelength number of Calcium ion and alginate molecule showed that crosslinked was occurred of all Caalginate microspheres (Figure 1-4).

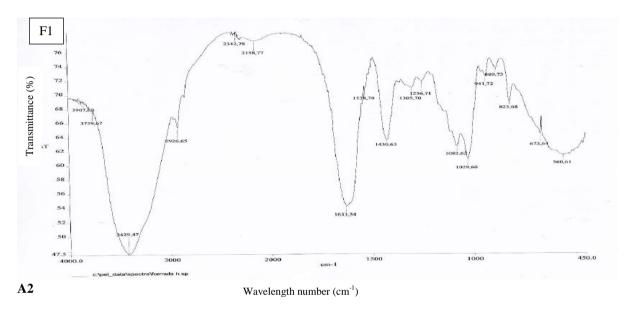


Figure 1. Infra Red spectra of formula A2

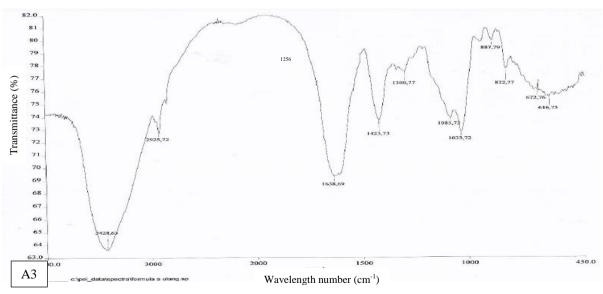


Figure 2. Infra Red spectra of formula A3

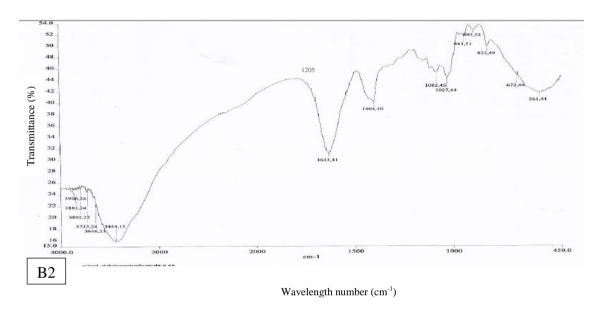


Figure 3. Infra Red spectra of formula B2

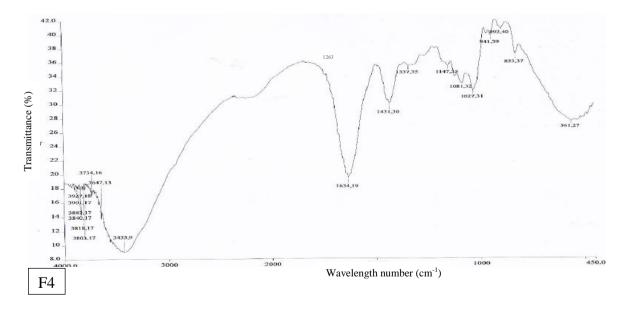


Figure 4. Infra Red spectra of formula B3

Three hundred particles of each formula were analyzed for particle size demonstrated average particle size of A2, A3, B1, B2 and B3 were 18,06 μ m; 12,54 μ m; 23,10 μ m; 21,14 μ m and 17,13 μ m respectively (**Figure 5**).

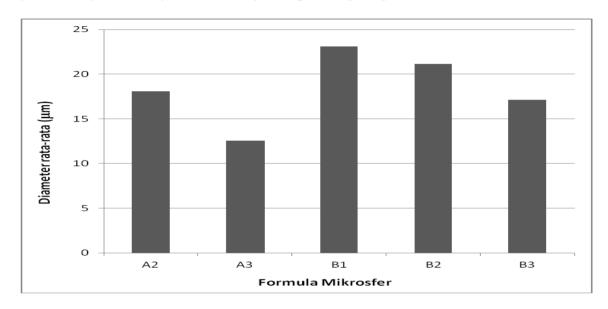


Figure 5. Particle size of ovalbumin – alginate microspheres from all formulas

Smaller particle size was shown by increasing concentration of CaCl₂, whereas by increasing alginate concentration at same concentration of CaCl₂, increased particle size. An increase of alginate concentration was due to an increase of alginate viscosity in forming bigger droplet which produced bigger microspheres'size (13). Additionally, current particle size was around 10 to 30 µm that also been demonstrated by Mishra et al (14) that immune response after oral administration could be achieved from microspheres with 1-30 µm in size. Manjanna et al (13) reported that by increasing concentration of Ca²⁺ formed spherical and smaller microsphere's size. This report was in agreement with Joshi *et al* (11) and Singh dan Kumar (15).

Formula	Appearance	Protein	Yield	Encapsulation
		Loading	(%)	Efficiency (EE)
		(%)		(%)
A1	Gelling sheet	-	-	-
A2	Microspheres	20.60 ± 5.78	33.56 ± 2.62	38.31 ± 10.38
A3	Microspheres	64.22 ± 18.82	62.30 ± 13.90	63.775 ± 4.33
B1	Microspheres	44.07 ± 13.17	50.43 ± 15.79	59.96 ± 28.92
B2	Microspheres	64,10 <u>+</u> 9,93	51,84 <u>+</u> 6,29	67,18 <u>+</u> 18,03
В3	Microspheres	74.50 ± 1.70	69.53 ± 5.01	88.80 ± 0.52

Encapsulation efficiency, protein loading and yield of microspheres can be seen in

TABLE 2. Encapsulation efficiency, protein loading and yield of microspheres

Table 2.

It was observed that larger amounts of CaCl₂ (from 0.1m to 1.5M), increased encapsulation efficiency ovalbumin in alginate microspheres (from 38% to 64% in formula A2 and A3; from 60% to 89% in formula B1 to B3). Significantly differences were found within formulas by statistic analysis. An increase of encapsulation efficiency is most likely caused by larger amounts of availability of Ca²⁺ that crosslinked with carboxylates from guluronic acid in alginate indicates more ovalbumin was entrapped within alginate microspheres (16). This trend was also similar to an increase of alginate concentration. The more alginate amounts, the more number of crosslinked alginate- CaCl₂, resulted the more ovalbumin was encapsulated (13). Similar studies were also confirmed that encapsulation efficiency increased by increasing concentration of polymer and crosslinking agents (11; 15).

The increase of protein loading from 20% to 64% in formula A2 and A3 as well as from 44% to 75% in formula B1 to B3 were most probably caused by an increase of concentration of CaCl₂, this may be again explained by strong network between carboxylates and Ca²⁺ ions in providing more space for ovalbumin inside microspheres (16).

Alginate microspheres produced using both alginate concentration (1.5 and 2.5%) using highest concentration of CaCl₂ (1.5M) indicated the highest yield of about 60% compare to formulas produced using lower concentration of CaCl₂. In the case of microspheres crosslinked using higher alginate concentration, the yield was also increased. This behaviour indicates that the more number of Ca²⁺ contact with alginate provide a gel network that able to increase yield of microspheres (10).

Conclusion

Ca-alginate microspheres entrapping model antigen Ovalbumin high entrapment efficiency, high protein loading, high yield and small particle size were successfully produced using this ionotropic gelation-aerosolisation method which can be utilized as one of oral protein or vaccine delivery systems.

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Declaration

The Authors declares that there is no conflict of interest.

References

O'neil, M. J., Patricia E. Heckelman, Cherie B. Koch, Kristin J. Roman, Catherine M. Kenny, Maryann R. D'Arecca. The Merck Index: An Encyclopedia Of Chemical Drug And Biological. 13th Ed., New Jersey: Merck & Co., inc. 2001.

- 2. Mowat, A., Mcl. The Role of antigen recognition and suppressor cells in mice with oral tolerance to ovalbumin. Immunology. 1985; 253-260.
- 3. Morris, W., Mark C., Philip,K. Potential of polymer microencapsulation technology for vaccine innovation. Vaccine, 1994; 12: 5-11.
- 4. Umer, H., Hemlata Nigam, Asif M Tamboli, M. Sundara Moorthi Nainar. Microencapsulation: Process, Techniques and Applications. International Journal of Research in Pharmaceutical and Biomedical Sciences. 2011; 2229-3701.
- 5. Yeo, Yoon, Baek N., Park, Kinam. Microencapsulation Methods for Delivery of Protein Drugs. Research Article, Biotechnology Bioinformatic Bioengineering. 2001: 213-230.
- 6. Dubey, M., Shami, T.C. and Rao, K.U. Bhasker. Microencapsulation Technology and Applications. Defence Science Journal, 2009; 59(1): 82-95.
- 7. Maria, M.S., Scher, Herbert, Jeoh, Tina. Microencapsulation of bioactives in cross-linked alginate matrice. Journal of Microencapsulation. 2012; 286-295.
- 8. Gombotz, W.R., Siow Fong Wee, Protein Release From Alginate Matrices. Advanced Drug Delivery Reviews, 1998; 31:267–285.
- 9. Putra, Joban. A.H., Bhushan Ashok Karode1, Sudhir Bhaskar Chincholkar. Calcium alginate as supporting material for the immobilization of rifamycin oxidase from Chryseobacterium species., Research Article, Biotechnology Bioinformatic Bioengineering. 2011; 529-535.
- 10. Jin, M., Yanping Zheng, Qiaohong Hu., Preparation and characterization of bovine serum albumin alginate/chitosan microspheres for oral administration. Asian Journal of Pharmaceutical Sciences. 2009; 215-220.
- 11. Joshi. S., Patel, P., Lin, S.Anda Madan, P.L. Development of Cross-Linked Alginate Spheres by Ionotropic Gelation Tecnique for Controlled Release of Naproxen Orally. Asian journal of Pharmacetical Science, 2012; 134 142.
- 12. Suksamran, Tittaya, Praneet Opanasopit, Theerasak Rojanarata, Tanasait Ngawhirunpat, Uracha Ruktanonchai dan Pitt Supaphol. Biodegradable alginate microparticles developed by electrohydrodynamic spraying techniques for oral delivery of protein. Journal of Microencapsulation, 2009; 563–570
- 13. Manjanna, K. M., Kumar, T. M. Pramod, B. Shivakumar. Calcium alginate cross-linked polymeric microbeads for oral sustained drug delivery in arthritis. Drug Discoveries & Therapeutics. 2010;109-122.
- 14. Mishra, Neeraj Amit K. Goyal, Kapil Khatri, Bhuvaneshwar Vaidya, Rishi Paliwal, Shivani Rai, Abhinav Mehta, Shailja Tiwari, Shiva Vyas, Vyas S. P., Biodegradable Polymer Based Particulate Carrier(s) for the Delivery of Protein and Peptides. Anti-Inflamatory & anti allergy Agent in Medical Chemistry. 2008; 240-251.
- 15. Singh, I. and Kumar, P., Formulation and optimization of tramadol loaded alginate beads using response surface methodology. Pak. J. Pharm. Sci, 2012; 25(4): 741-749
- 16. Gulati, Neha, Upendra Nagaich, V.K Sharma, R.L Khosa. Effect of Polymer and Cross Linking Agent on In Vitro Release of Quercetin from Microbeads. Asian Journal of Pharmacy and Life Science. 2011; 1-5.